



Research paper

Cell-penetrating conjugates of pentaglutamylated methotrexate as potential anticancer drugs against resistant tumor cells

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ABSTRACT

The emerging resistance of tumor cells against methotrexate (MTX) is one of the major limitations of the MTX treatment of tumorous diseases. The disturbance in the polyglutamation which is a main step in the mechanism of methotrexate action is often the reason of the resistance. Delivery of polyglutamylated MTX into cells may evade the mechanisms that are responsible for drug resistance. In this study conjugates of methotrexate and its pentaglutamylated derivatives with cell-penetrating peptides – penetratin and octaarginine – were investigated. The cellular-uptake and *in vitro* cytostatic activity of conjugates were examined on breast cancer cell cultures (MDA-MB-231 as resistant and MCF-7 as sensitive cell culture). These cell cultures showed very different behaviour towards the conjugates. Although the presence of pentaglutamyl moiety significantly decreased the internalisation of conjugates, some of them were significantly active *in vitro*. All of the conjugates were able to penetrate in some extent into both cell types, but only the conjugates of penetratin showed *in vitro* cytostatic activity. The most effective conjugates were the MTX-Glu₅-Penetratin(desMet) and MTX-Glu₅-GFLG-Penetratin(desMet). The latter was effective on both cell cultures while the former was active only on the resistant tumor cells. Our results suggest that the translocation of polyglutamylated MTX may be a new way to treat sensitive and more importantly resistant tumors. While both penetratin and octaarginine peptides were successfully used to deliver several kinds of cargos earlier in our case the activity of penetratin conjugates was more pronounced.

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1. Introduction

Methotrexate (4-amino-10-methylfolic acid, MTX) was the first antimetabolite used in the treatment of tumorous diseases from the 1950s. As an effective anticancer drug it is clinically used to treat childhood acute lymphoblastic leukemia and a number of other haematologic malignancies [1]. Methotrexate, belonging to the antifolate antimetabolite family, can be internalised into cells by specific transporters - the membrane-bound reduced folate carrier (RFC-1) and/or folate-binding proteins (folate receptors Fr α , FR β and FR γ) [2]. Inside the cell sequential addition of Glu moiety/moieties occurs by the enzyme folylpolyglutamate synthetase (FPGS) [3–5]. Polyglutamylated MTX exhibiting higher affinity to

folate-metabolizing enzymes play crucial role in the metabolism of folic acid and disturb the biosynthesis of thymidine and purine, inhibit the synthesis of DNA and could lead to cell death.

During the polyglutamation 5–8 glutamic acid residues are coupled to the MTX *via* amide linkage forming γ -peptide bonds. The MTX is a poor substrate of the FPGS which catalyses the glutamic acid attachment and its binding competes with intracellular folates [6]. The polyglutamylated forms of MTX can inhibit the dihydrofolate reductase and thymidylate synthase enzymes. The presence of the polyglutamyl moiety essentially prevents the efflux of MTX from the cell because derivatives containing more than three glutamic acid residues are not substrates of multidrug resistance protein (MRP 3) [7] and of the folate transporter systems [8]. Thus the polyglutamation is a very important process in the action of MTX. The concentration of polyglutamylated MTX in the cytosol is modulated by the activity of FPGS and of the enzyme γ -glutamyl hydrolase (GGH), catalysing the removal of Glu residues [9,10]. Therefore, the presence of polyglutamylated MTX derivatives depends on the balance of glutamic acid attachment and removal.

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Although several second and third generation antifolates have been developed for clinical use, the main drawback is the development of the evolving drug resistance. There are various processes which are responsible for the drug resistance: i) decreased cellular uptake caused by the malfunction of folate transporters [11]; ii) increased efflux related to the overexpression of multidrug resistance transporters [12]; and/or iii) decreased level of polyglutamation due to inactivation or decreased activation of respective enzymes that take part in these processes [13]. The drug resistance can be intrinsic or acquired. Some studies showed that the defect in the polyglutamation can result in the intrinsic drug resistance [14,15]. Therefore the intracellular delivery of polyglutamylated MTX may provide an effective antitumour drug against resistant tumor cells as well.

Polyglutamylated MTX derivatives are very hydrophilic, and they are not substrates of folate transporter systems (RFC-1 and/or FRs) delivering folates into the cells. Consequently, these compounds are not able to enter through the cell membrane. This disadvantage may be overcome by using cell-penetrating peptides (CPPs). CPPs are short oligopeptides (natural and/or of synthetic origin) with cell internalisation capability. Among the first peptides discovered, derivatives from proteins: HIV-1 Tat [16,17] and Antennapedia protein of *Drosophila* could be mentioned [18]. These oligopeptides are able to penetrate across the cell membrane even in the presence of various, covalently attached cargos (e.g. peptides, peptide-nucleic acids, oligonucleotides) [19].

For targeted delivery into sensitive and/or resistant tumor cells MTX or pemetrexed [20] has been conjugated to protein [21], peptides including CPP [22–25], polymeric polypeptides [26], dendrimers [27,28] and oligosaccharides [29].

In spite of the potency of polyglutamylated MTX as effective drug against resistant tumors only few studies were reported on conjugates with this molecule. An early paper described conjugates in which MTX with different length of polyglutamyl chains were attached to polymeric polylysine (average $M = 27,000$ and $52,000$) [30]. In these conjugates the polyglutamylated MTX was coupled *via* amide bond between the α/γ -carboxylic group of glutamic acids and ϵ -amino group of lysine side chains. Since the unprotected form was used in the conjugation reaction the exact structure of the conjugates (the carboxylic group involved in the formation of the amide bond) is unknown. The number of MTX attached to the polymer was between 2 and 11. The conjugates showed inhibitory activity of sensitive (H35) and MTX transport resistant (H35R) strains of hepatoma cells. In a recent study the polyglutamylated forms of MTX were conjugated with sequence-defined oligo (ethanamino)amide [31]. These compounds and the free MTX exhibited similar activity against human cervix carcinoma KB cells *in vitro*.

Based on our promising results on cell penetrating peptide - antitumour drug (daunomycin, vinblastine [32,33]) or enzyme activator/substrate peptide [34,35] conjugates our aim was to study the cell-penetrating peptide concept to deliver pentaglutamylated methotrexate into tumor cells. Here we describe the synthesis and characterization of a novel set of conjugates in which pentaglutamylated MTX is covalently attached to the N-terminal of cell-penetrating peptides *via* peptide bond. Two conjugate families are reported here with two different types of CPPs (octaarginine or modified penetratin) possessing different mechanisms of uptake. The cytostatic effect as well as cellular uptake properties of conjugates were studied on sensitive and resistant breast tumor cell lines *in vitro*. Data collected were comparatively analysed with those obtained with MTX-conjugates without pentaglutamyl moiety as well. We observed that the presence of penetratin (desMet) highly influenced the activity of pentaglutamylated MTX on resistant breast cancer cells even in comparison with the MTX-

conjugate, but not on MCF-7 cells. Interestingly, no *in vitro* cytotoxic effect was documented with conjugate containing Arg₈ as CPP. Our results show that the insertion of an enzyme labile spacer (GFLG) between pentaglutamylated MTX and penetratin (desMet) could maintain (MDA-MB-231) or increase (MCF-7 cell) the activity of the conjugate as well.

2. Materials and methods

All amino acid derivatives, *N,N'*-diisopropylcarbodiimide (DIC) and Rink-amide MBHA resin were purchased from IRIS Biotech GmbH (Marktredwitz, Germany). *N,N'*-diisopropylethylamine (DIEA), 1,8-diazabicyclo [5.4.0]undec-7-ene (DBU), thioanisole, 1,2-ethanedithiol (EDT) were FLUKA (Buchs, Switzerland), while 1-hydroxybenzotriazole (HOBt), trifluoroacetic acid (TFA) and phenol were Sigma Aldrich (Budapest, Hungary) products. Solvents for synthesis and purification were obtained from Molar Chemicals Ltd (Budapest, Hungary). Carboxyfluorescein (cf) and all other chemicals used in biological experiments were purchased from Sigma Aldrich (Hungary). Buffers were prepared with distilled water.

2.1. Synthesis of peptides and peptide-conjugates

The conjugates were built up by solid-phase peptide synthesis on Rink amide MBHA resin using Fmoc/^tBu strategy. The side chain of amino acids were protected by 2,2,4,6,7-pentamethylidihydrobenzofuran-5-sulfonyl group (Pbf, in Arg), by trityl (Trt, in Asn, Gln), by *tert*-butyl group (^tBu, in Glu) and by *tert*-butyloxycarbonyl group (Boc, in Lys). The N-terminal Fmoc protecting group was removed with 2% piperidine in the presence of 2% 1,8-diazabicyclo [5.4.0]undec-7-ene (DBU) in DMF (2 + 2+5 + 10 min). This reagent was removed by washing with DMF (8 × 0.5 min). In the coupling reaction, amino acid derivatives, which were used in 3 M excess to the resin capacity, were activated by *N,N'*-diisopropylcarbodiimide (DIC) and 1-hydroxybenzotriazole (HOBt) dissolved in DMF. The reaction proceeded at RT for 60 min. Then, the resin was washed with DMF (2 × 0.5 min) and dichloromethane (DCM) (3 × 0.5 min). Ninhydrin assay was used to monitor the efficiency of the coupling reaction [36]. After the removal of the terminal *N*^α-Fmoc group, methotrexate (MTX) or 5 (6)-carboxyfluorescein (cf) were attached to the peptide using the HOBt-DIC coupling reagents. The free compounds were obtained by cleavage from the resin with 10 mL TFA containing 0.75 g phenol, 0.5 mL distilled water, 0.5 mL thioanisole and 0.25 mL 1,2-ethanedithiol (EDT) as scavengers. Crude products were precipitated by dry diethyl-ether, dissolved in 10% acetic acid and freeze-dried. The crude peptide was purified by RP-HPLC as described below. The purified compounds were characterized by analytical RP-HPLC and ESI-MS (Table 1). The purity of the compounds was higher than 95%.

2.2. RP-HPLC

Analytical RP-HPLC was performed on a Knauer (Herbert Knauer GmbH, Berlin, Germany) HPLC system using a Phenomenex Jupiter C18 column (250 × 4.6 mm I.D.) with 5 μm silica (300 Å pore size) (Torrance, CA USA) as a stationary phase. Linear gradient elution (0 min 0% B; 5 min 0% B; 50 min 90% B) with eluent A (0.1% TFA in water) and eluent B (0.1% TFA in acetonitrile-water (80:20, V/V)) was used at a flow rate of 1 mL/min at ambient temperature. Peaks were detected at λ = 220 nm. The samples were dissolved in eluent B. In the other cases Exformma (Exformma Technology (ASIA) Co., Ltd, Hong Kong, China) HPLC system was used. The column was Agilent Zorbax SB-C18 4.6 mm × 150 mm, 100 Å. The applied linear

Table 1
Chemical characteristics of peptides and peptide-conjugates.

| Compounds | Rt (min) | ESI-MS ^c | |
|---|-------------------|---------------------|--------|
| | | Mcal. | Mmeas. |
| Arg ₈ ^a | 10.3 ^b | 1266.5 | 1266.3 |
| Penetratin ^a | 12.9 ^b | 2245.7 | 2246.2 |
| Mtx-Arg ₈ | 21.2 ^a | 1702.6 | 1703.7 |
| Mtx-Penetratin(desMet) ^a | 24.5 ^a | 2251.0 | 2251.4 |
| Mtx-Glu ₅ -Arg ₈ | 21.9 ^a | 2348.5 | 2349.0 |
| Mtx-Glu ₅ -Penetratin(desMet) | 25.3 ^a | 3196.6 | 3197.1 |
| Mtx-GFLG-Arg ₈ | 12.6 ^b | 2077.4 | 2077.8 |
| Mtx-GFLG-Penetratin(desMet) | 14.1 ^b | 2925.4 | 2925.9 |
| Mtx-Glu ₅ -GFLG-Arg ₈ | 12.4 ^b | 2722.9 | 2723.3 |
| Mtx-Glu ₅ -GFLG-Penetratin(desMet) | 14.1 ^b | 3571.0 | 3570.9 |
| cf-Arg ₈ | 11.6 ^b | 1624.8 | 1625.4 |
| cf-Penetratin(desMet) | 14.1 ^b | 2472.9 | 2473.2 |
| cf-Glu ₅ -Arg ₈ | 22.9 ^a | 2270.4 | 2270.7 |
| cf-Glu ₅ -Penetratin(desMet) | 27.8 ^a | 3118.4 | 3119.4 |
| cf-GFLG-Penetratin(desMet) | 15.9 ^b | 2847.3 | 2846.9 |
| cf-Glu ₅ -GFLG-Penetratin(desMet) | 15.0 ^b | 3492.9 | 3492.8 |

^a Penetratin: RQIKIWFQNRRMKWKK, Arg₈: RRRRRRRR, Penetratin(desMet): RQIKIWFQNRRMKWKK. The purity of the compounds was higher than 95 %.

^b RP-HPLC retention time value. Column: Phenomenex Luna C18(2) 5 μM, 250 × 4.6 mm, 100 Å. Linear gradient elution: 0 min 0% B; 5 min 0% B; 50 min 90% B with eluent A (0.1% TFA in water) and eluent B (0.1% TFA in acetonitrile-water (80:20, V/V)) were used at a flow rate of 1 mL/min at ambient temperature. Peaks were detected at λ = 220 nm.

^c Column: Agilent Zorbax SB-C18 4.6 × 150 mm, 100 Å; Gradient: 0 min 0% B, 2 min 0% B, 22 min 90% B; Flow rate: 1 mL/min; RT, λ = 220 nm.

^d ESI-MS analysis was carried out on a Bruker Esquire 3000 plus equipment (Germany). The sample was dissolved in acetonitrile-water (50:50, V/V), 0.1% acetic acid.

gradient elution was 0 min 0% B, 2 min 0% B, 22 min 90% B with 1 mL/min flow rate. The detection was carried on at λ = 220 nm.

The crude products were purified on a semi-preparative Phenomenex Jupiter C18 column (250 × 10 mm I.D.) with 10 μm silica (300 Å pore size) (Torrance, CA, USA). Flow rate was 4 mL/min. Linear gradient elution was applied. The samples for purification were dissolved in eluent A containing few percent eluent B.

2.3. Mass spectrometry

The molecular mass of peptides as well as conjugates was determined by ESI-MS. Electrospray ionization mass spectrometric analysis was performed on a Bruker Daltonics Esquire 3000 plus (Germany) ion trap mass spectrometer. The samples were dissolved in acetonitrile–water (50:50, V/V), containing 0.1% acetic acid. Samples were analysed by direct injection using a syringe pump. Instrumental parameters were the following: capillary voltage: 4 kV, nebulizer gas: 10 psi, dry gas: 4 L/min, heated capillary temperature: 250 °C.

2.4. Cell cultures for *in vitro* studies

MCF-7 (ATCC: HTB-22) human breast adenocarcinoma [37] and MDA-MB-231 (ATCC: HTB-26) human triple negative breast adenocarcinoma [38] cells were used for the analysis of the *in vitro* cytostatic effect of MTX containing conjugates and controls (MTX, Arg₈ and penetratin) and also for the investigation of the cellular uptake of carboxyfluorescein labelled compounds. MCF-7 cells were maintained in DMEM supplemented with 10% heat-inactivated foetal calf serum (FCS), non-essential amino acids (NEAA), pyruvate (1 mM), L-Gln (2 mM) and gentamicin (160 μg/mL). MDA-MB-231 cells were cultured in RPMI-1640 supplemented with 10% FCS, L-Gln (2 mM) and gentamicin (16 μg/mL). Cells were maintained in plastic tissue culture dishes at 37 °C with a humidified atmosphere containing 5% CO₂/95% air.

2.5. Determination of the *in vitro* cellular uptake profile by flow cytometry

MCF-7 and MDA-MB-231 cells were cultured as described above. In order to study the cellular uptake of fluorescently labelled compounds, 10⁵ cells per well were plated on 24-well plates. After 24 h incubation at 37 °C, cells were treated with the compounds in serum-free medium for 90 min. The cellular uptake was analysed at 1, 5 and 10 μM concentrations. Cells treated with serum-free medium for 90 min were used as negative control. After incubation, the solutions of conjugates were removed from the cells and the cells were treated with 100 μL trypsin for 10 min. The activity of trypsin was terminated by the addition of 900 μL HPMI (glucose, NaHCO₃, NaCl, HEPES, KCl, MgCl₂, CaCl₂, Na₂HPO₄·2H₂O) containing 10% foetal calf serum, and the cells were moved from the plate to FACS-tubes. Cells were centrifuged at 216 g at 4 °C for 5 min and the supernatant was removed. Cells were re-suspended in 500 μL HPMI. The fluorescence intensity of cells was monitored by flow cytometry (BD LSR II, BD Bioscience, San Jose, CA). Data were analysed with FACSDiVa 5.0 software.

2.6. Analysis of the *in vitro* cytostatic activity of the conjugates

The cells were grown to confluency and were plated into 96-well plate with initial cell number of 5·10³ per well. After 24 h incubation at 37 °C, cells were treated with the compounds at 1.28·10⁻³–100 μM concentration range for 3 h in 200 μL final volume. Control cells were treated with serum free medium at 37 °C for 3 h. After incubation the cells were washed twice with serum free medium. For the analysis of the *in vitro* cytostatic effect, cells were cultured for an additional 72 h in serum containing medium. On day 4, MTT assay [39,40] was carried out in order to determine the IC₅₀ values of the compounds. Briefly, 45 μL of MTT solution was added to each well (2 mg/mL, dissolved in serum-free medium). Following the 4 h incubation, plates were centrifuged at 900 g for 5 min, and the supernatant was removed. The precipitated purple crystals were dissolved in 100 μL DMSO and the absorbance was determined at λ = 540 nm and 620 nm using ELISA plate reader (iEMS Reader, Labsystems, Finland). The percent of cytostasis was calculated using the following equation: Cytostatic effect (%) = [1 – (OD_{treated}/OD_{control})] × 100; where OD_{treated} and OD_{control} correspond to the optical densities of the treated and the untreated control cells, respectively. In each case three independent experiments were carried out with 4 parallel measurements. The 50% inhibitory concentration (IC₅₀) values were determined from the dose-response curves. The curves were defined using MicrocalTM Origin (version 9.2) software: cytostasis was plotted as a function of concentration, fitted to a sigmoidal curve, and based on this curve, the IC₅₀ value was determined. IC₅₀ represents the concentration of a compound required for 50% inhibition *in vitro* and expressed as micromolar units.

3. Results

After the internalisation MTX typically undergoes polyglutamation [3–5]. These derivatives (MTX-(Glu)_n where n = 5 to 8) are very important to maintain the effective intracellular concentration. Alteration of the homeostasis of the polyglutamylated MTX forms often results in the development of drug resistance [13]. For this study we have selected the pentaglutamylated derivative (MTX-Glu₅) considering that this compound is not a substrate of folate transport systems. We have examined the effect of the pentaglutamylated derivative (MTX-Glu₅) on the cell viability as well as on the uptake properties of two different tumor cell cultures. For intracellular delivery its conjugates with cell-penetrating

peptides were also synthesised, since - due to its negative charges - this methotrexate derivative is not able to penetrate across the cell-membrane.

3.1. Synthesis of the conjugates

The different cell-penetrating peptides could have very diverse internalisation mechanisms and thus have high influence on the intracellular fate of the covalently attached cargo moiety. Although both octaarginine and penetratin are positively charged with several arginine residues in the sequence, based on the characteristics of their internalisation, the two peptides belong to different classes of cell-penetrating peptides [41]; penetratin is secondary amphipathic [42], while octaarginine is non-amphipathic cell-penetrating peptide [43]. For studying whether the difference between the two CPP's has any effect on *in vitro* cytostatic activity, MTX-conjugates of both peptides were prepared. Since methionine in the penetratin sequence could be oxidized under various conditions and might decrease the stability of conjugates, penetratin derivative without methionine (RQIKIWFQNRRKWK (Penetratin (desMet))) was produced. Earlier studies [44] as well as our own experience (data is not shown) proved that this Met residue can be removed from the sequence without decreasing the internalisation ability.

During the intracellular attachment of polyglutamyl chain γ -peptide bond formed between the glutamic acids. However, the cell has also an apparatus to remove this chain from MTX [9,10]. Therefore to avoid the decomposition of our MTX derivative by gamma-glutamyl hydrolase, the glutamic acid residues were connected with their α -carboxyl group.

The conjugates of MTX and MTX-Glu₅ (Fig. 1) with cell-penetrating peptide were synthesised by solid phase peptide synthesis using standard Fmoc/^tBu strategy.

First the cell-penetrating peptides with or without pentaglutamyl moiety at their N-terminus were built up and MTX was attached to the free N-terminal amino group of peptides using DIC and HOBt coupling reagents. Then the conjugates were cleaved from the resin by TFA cleavage mixture and were purified by RP-HPLC as described in the Experimental section. The characterisation of conjugates was performed by analytical RP-HPLC and ESI-

MS (Table. 1). During the purification only one peak was isolated with the correct molecular mass. As the MTX contains two carboxyl groups and none of them was protected, the formation of two main isomers could be expected during the synthesis of MTX-conjugates. Besides this the racemisation of glutamic acid residue in the MTX molecule may also occur, the D-isomers might be also present in the mixture. Based on earlier results using the same coupling strategy but different peptide component, MTX mainly coupled via its γ -carboxyl group in these circumstances [22]. Thus we could isolate the mixture of isomer conjugates containing mainly γ -peptide bond between the MTX and the peptides. For the cellular uptake studies fluorescence labelled conjugates were also prepared. These derivatives were synthesised as MTX conjugates, but cf was attached instead of MTX.

Since we know little about the fate of internalised conjugates, and about the potential release of MTX and MTX-Glu₅, we have prepared conjugates with a tetrapeptide spacer (GFLG) cleavable by a lysosomal enzyme cathepsin B inserted between the cell-penetrating peptide and drug component. The cytostatic as well as uptake properties of these compounds were also studied.

3.2. *In vitro* cytostatic effect of the conjugates

The *in vitro* cytostatic activity of MTX-conjugates was examined on two tumor cell lines, MCF-7 human breast adenocarcinoma and MDA-MB-231 human triple negative breast adenocarcinoma cell lines (Table. 2). The main difference between these two cell cultures is their receptor status. MCF-7 cells express oestrogen and progesterone receptors, but MDA-MB-231 cells are oestrogen, progesterone and HER2 receptor negative (called triple negative) cells [45]. This different receptor status could cause significant differences in the therapeutic responses to various drugs, like in case of MTX. Considering that MCF-7 cells are sensitive to MTX treatment, while MDA-MB-231 cells are MTX resistant, these two cell cultures were selected as models for our *in vitro* experiments. It is important to note that the drug resistance of MDA-MB-231 cells is due to the lack of reduced folate carrier [46].

On MCF-7 cells the free MTX was highly active with IC₅₀ = 0.8 μ M, while the free cell-penetrating peptides, as expected exhibited no activity in the concentration range studied here

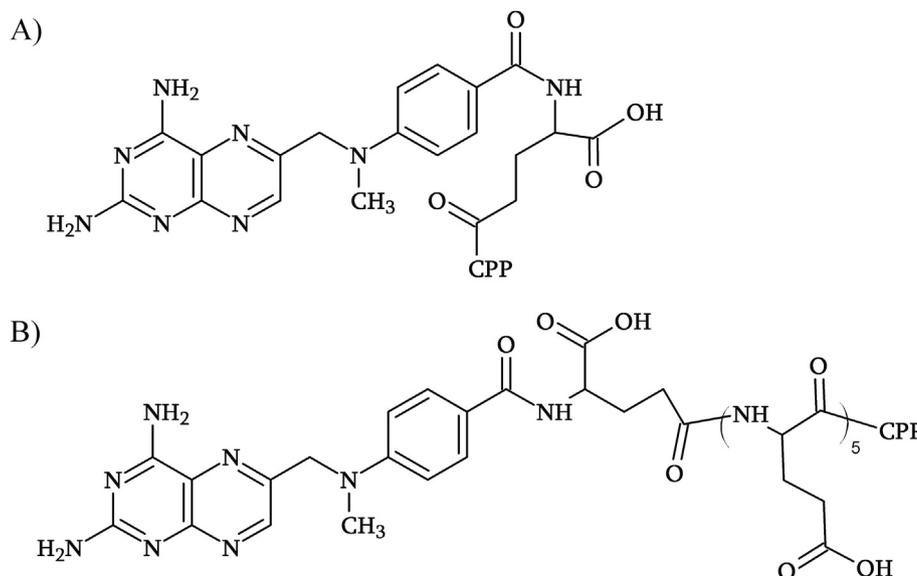


Fig. 1. The structure of MTX (A) and MTX-Glu₅ (B) containing CPP-conjugates.

Table 2*In vitro* cytostatic activity of MTX-conjugates on tumor cells.

| Compound | IC ₅₀ (sd) (μM) ^a | |
|---|---|-------------|
| | MDA-MB-231 | MCF-7 |
| Mtx | > 100 | 0.8 (0.3) |
| Arg ₈ | > 100 | > 100 |
| Penetratin | > 100 | > 100 |
| Mtx-Arg ₈ | ~ 100 | > 100 |
| Mtx-Penetratin(desMet) | 12.9 (4.0) | 40.2 (14.4) |
| Mtx-Glu ₅ -Arg ₈ | > 100 | > 100 |
| Mtx-Glu ₅ -Penetratin(desMet) | 0.1 (0.1) | >100 |
| Mtx-GFLG-Arg ₈ | 19.1 (3.7) | 31.3(9.6) |
| Mtx-GFLG-Penetratin(desMet) | 5.5 (0.5) | 13.9 (0.5) |
| Mtx-Glu ₅ -GFLG-Arg ₈ | ~100 | > 100 |
| Mtx-Glu ₅ -GFLG-Penetratin(desMet) | 0.4 (0.1) | 0.3 (0.1) |

^a The cells were incubated with the compound for 3 h, after cultured in serum-containing medium for 3 days. The IC₅₀ values were determined by MTT assay as described in the text. Standard deviation values (sd) are also presented.

(IC₅₀ > 100 μM). Among the two MTX conjugates only the MTX-Penetratin (desMet) showed some moderate activity (IC₅₀ = 40.2 μM), while the octaarginine conjugate was completely inefficient. The conjugates of MTX-Glu₅ were fully inactive. The influence of enzyme labile spacer (GFLG) was also studied. The presence of the GFLG tetrapeptide spacer both in the Arg₈ and penetratin (desMet) conjugates of MTX increased the activity (IC₅₀ = 31.3 μM for MTX-GFLG-Arg₈ and IC₅₀ = 13.9 μM for MTX-GFLG-Penetratin (desMet)). In case of MTX-Glu₅ conjugates the presence of the spacer resulted in higher activity only in case of MTX-Glu₅-GFLG-Penetratin (desMet) (IC₅₀ = 0.3 μM), while the octaarginine conjugate was ineffective.

In the agreement with the literature [46], free MTX was ineffective against the resistant MDA-MB-231 cells (IC₅₀ > 100 μM) as compared to the MCF-7 cells (Table 2). The cell-penetrating peptides had also no effect on these cells in the applied concentration range (IC₅₀ > 100 μM). Although the cell-penetrating peptide can transport the covalently attached MTX very effectively (see the next section), only the MTX-Penetratin (desMet) had low activity (IC₅₀ = 12.9 μM). Incorporation of the pentapeptide of Glu resulted in a highly cytostatic compound (IC₅₀ = 0.1 μM). In contrast, similar conjugate with octaarginine did not improve the cytostatic effect against the resistant MDA-MB-231 cells (IC₅₀ = 100 μM for both MTX-Arg₈ and MTX-Glu₅-Arg₈). Incorporation of the GFLG tetrapeptide spacer between the cell-penetrating peptide and MTX increased the activity of both conjugates. The presence of this tetrapeptide did not elevate the activity of MTX-Glu₅ conjugates. Based on the *in vitro* cytostatic effect on MDA-MB-231 cells the following order could be established among conjugates containing penetratin (desMet): MTX-Glu₅-Penetratin (desMet) (IC₅₀ = 0.1 μM) > MTX-Glu₅-GFLG-Penetratin (desMet) (IC₅₀ = 0.4 μM) > MTX-GFLG-Penetratin (desMet) (IC₅₀ = 5.5 μM) > MTX-Penetratin (desMet) (IC₅₀ = 12.9 μM).

3.3. Internalisation of the conjugates

Based on the *in vitro* cytostatic activity of the conjugates, cellular uptake profile of some conjugates was investigated by flow cytometry. Considering difficulties in measuring the cellular uptake based on the spectroscopic properties of MTX we have prepared conjugates in which the drug was replaced by 5 (6)-carboxy-fluorescein (cf). The cellular uptake profile of these compounds was studied and accepted as an approximation for the uptake of the relevant MTX-Glu₅-CPP or MTX-CPP conjugates.

First, we were interested in studying the uptake properties of conjugates containing five glutamic acid residues by MTX sensitive

and resistant cells. MCF-7 and MDA-MB-231 cells were treated with the solution of cf labelled compounds using in the concentration range of 1–10 μM at RT for 90 min. After washing and trypsin treatment the fluorescence intensity of the cells was measured by flow cytometry (Fig. 2). All conjugates were able to penetrate into the cells and the amount of internalised conjugates was increased by the elevated concentration. Based on the fluorescence intensity of the cells the internalisation of cf labelled octaarginine was higher than that of cf-Penetratin (desMet) on both cell lines. The intracellular amount of octaarginine was significantly increased by the concentration (mean fluorescence of MCF-7 cells: 975 (1 μM) < 8669 (5 μM) < 52,928 (10 μM), Fig. 2A). The cellular-uptake of penetratin (desMet) also showed growing value with the concentrations, but the rise was lower (mean fluorescence of MCF-7 cells: 1728 (1 μM) < 7361 (5 μM) < 12,310 (10 μM), Fig. 2A). The internalisation of octaarginine was more pronounced on MDA-MB-231 cells while the penetratin had similar cellular uptake on both cell lines.

The presence of pentaglutamyl moiety significantly reduced the

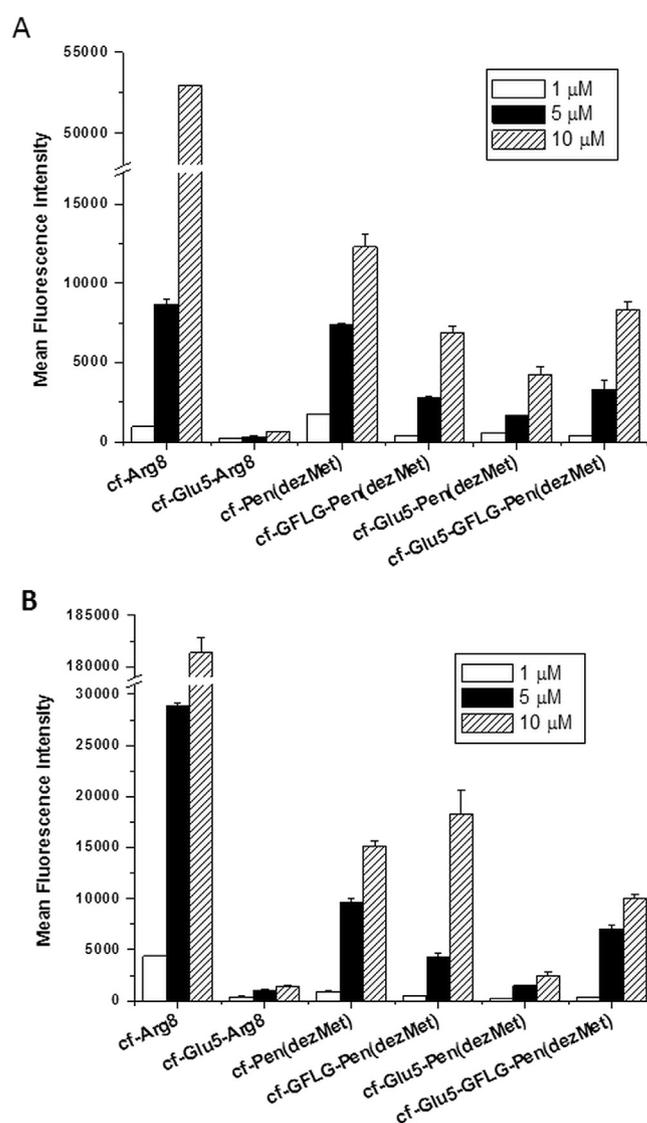


Fig. 2. Cellular-uptake profile of fluorescently labelled conjugates by different cells A) MCF-7 and B) MDA-MB-231 cells were treated with the solution of the conjugates (1, 5 and 10 μM) for 90 min. The fluorescence intensity of the cells was determined by flow cytometry after trypsin treatment.

cellular uptake of the cell-penetrating peptides. This reduction was the highest in case of octaarginine on both cell cultures. The fluorescence intensity decreased below the 10% of the non-glutamylated form. In case of penetratin (desMet) the reduction was lower and the pentaglutamylated penetratin (desMet) could be internalised more efficiently than the octaarginine derivative. Not only the fluorescence intensity, but also the number of fluorescence positive cells was decreased. At 1 μM concentration only a few percent of the cells engulfed the pentaglutamylated conjugates (data is not shown).

We also studied the effect of tetrapeptide spacer on the internalisation of penetratin (desMet) derivatives. Incorporation of GFLG between the pentaglutamyl moiety and the penetratin (desMet) increased significantly the cellular uptake, but it was still lower in comparison with the free penetratin (desMet).

4. Discussion

The MTX is an active clinically used antitumour drug. Unfortunately tumor cells may become resistant against it due to the development of several potential processes. Two of these are in relation with the inadequate accumulation of MTX in the cytosol caused by the decreased influx or increased efflux and the lack of polyglutamation. In case of sensitive cells the MTX influx is adequate to achieve the proper intracellular concentration and the drug could inhibit the proliferation (for example MCF-7 cells; Table. 2). If this influx is insufficient drug resistance could develop (for example MDA-MB-231 cells; Table. 2). The accumulation of MTX in sensitive or resistant cells can be enhanced for example by using delivery vehicle (peptides, polymers, nanoparticles) or in the absence of polyglutamation mechanism by intracellular targeting of polyglutamylated MTX. Among others, two main steps could influence the activity of free and conjugated MTX derivative: internalisation across the cell-membrane, and the interaction with the target enzymes.

In our systematic study we compared the *in vitro* antitumour effect of new cell-penetrating conjugates of MTX and MTX-Glu₅. Two cell-penetrating peptides were applied in these conjugates, octaarginine and penetratin without Met (penetratin (desMet), with or without tetrapeptide spacer (GFLG). From these compounds MTX might be released only after the degradation of peptide and/or the enzymatic cleavage of GFLG spacer. As the GFLG spacer can be cleaved by Cathepsin B lysosomal enzyme both the degradation and the enzyme cleavage can occur in case of vesicular transport, but not in case of direct translocation. Therefore the activity of the MTX and MTX-Glu₅ conjugates mainly depends on the internalisation properties and inhibitory activity of the conjugates.

The *in vitro* cytostatic effect of the conjugates was measured on resistant and sensitive breast cancer cell lines; MDA-MB-231 and MCF-7 cells, respectively. The MTX was active only on MCF-7 cells and did not show any activity on the resistant cell culture. The effect of cell-penetrating peptides on the cytostatic activity was very divergent. Only the penetratin derivatives exhibited activity. Although the activity of this conjugate was lower on MCF-7 cells compared to the free MTX, it was effective against MDA-MB-231 cells too. The reduction of the efficiency of MTX in the conjugates could be explained by the limited inhibition of the target enzymes by the conjugated forms. Earlier studies reported that conjugation with different carrier molecules decreased the enzyme inhibitory activity of MTX [25,31]. The alteration of the efficacy of octaarginine and penetratin (desMet) conjugates cannot be explained with the different internalisation ability of the cell-penetrating peptides. However, octaarginine efficiently penetrate into both cell lines (Fig. 2), it has no *in vitro* cytostatic activity (Table.

2). According to this observation, it was supposed that the internalised octaarginine conjugates cannot inhibit the target intracellular enzymes as much as the penetratin (desMet) ones. For studying if the mechanism of internalisation may cause this difference, enzyme labile spacer was introduced between the MTX and the CPP. The GFLG tetrapeptide spacer applied can be cleaved by cathepsin B enzyme in the lysosomes. Thus if the cells take up the conjugates by any kind of vesicular transport, the cleavage of this spacer may enhance the activity of the compound. While the presence of this spacer slightly increased the effect of penetratin (desMet) conjugate, it is significantly increased the activity of octaarginine conjugate. Based on these findings we suppose that a) the octaarginine conjugate mainly penetrate into the cells by vesicular transport, which is accordance with published data about the internalisation of oligoarginines [47] and/or b) the release of MTX from the conjugate is necessary.

In case of the conjugates of MTX-Glu₅, the octaarginine containing one had again no activity on MCF-7 and MDA-MB-231 cells. The penetratin (desMet) based conjugate was very active on resistant cells, but was inactive on sensitive cells. The effect of this conjugate was higher than that of the free MTX on sensitive cells. The presence of pentaglutamyl moiety in the conjugates reduces dramatically the internalisation of CPP's on both cell lines. This reduction was more pronounced in case of the octaarginine construct. Our data are in accord with earlier data describing that the positively charged guanidine groups are essential in the internalisation of oligoarginines [48]. The negatively charged glutamic acid residues which could be close to the positively charged guanidine groups can establish electrostatic interaction and thus prevent the binding of cell-penetrating peptide to the cell membrane. This interpretation could be supported with the class of "activatable cell-penetrating peptides" [49]. In these constructs poly anionic peptide domains (mainly Glu_n) can inhibit the internalisation of poly cationic oligoarginines *via* electrostatic interaction. Removal of this inhibitory domain results in increased cellular uptake. The results of the cellular-uptake study published [50] showed that the arginines are important in the internalisation of penetratin (desMet), but their possible blocking in our constructs do not diminish fully the uptake. The effect of penetratin (desMet) conjugates on viability was more marked in the case of resistant MDA-MB-231 cells. While penetratin (desMet) can internalise at the similar extent into both cells, the cellular-uptake of pentaglutamylated derivative by resistant cells was smaller. The difference between the *in vitro* cytostatic activities of the two sets of conjugates may be explained with the altered mechanism of cellular uptake and/or with the altered enzyme sets present in the two cell lines. Based on the fluorescence intensity of the cells only the internalised amount of conjugates was measured, but the mechanism of the penetratin (desMet) translocation needs further investigations, considering that the mechanism of uptake can influence the intracellular distribution of the conjugates (cytosol vs vesicles) and thus the interactions with intracellular target enzymes. The altered mechanism may be responsible for the divergent *in vitro* cytostatic activity. However, to best of our knowledge there are no data in the literature about the cell-line dependent internalisation mechanism of cell-penetrating peptides. The other possible explanation of differences may be due to the altered enzyme set of different cells, and the differential binding ability of conjugates to these enzymes.

The unwanted interaction of pentaglutamyl moiety with the positively charged cell-penetrating peptides may cause the ineffectiveness of conjugates. Therefore the effect of spacer was also studied. This spacer increases the distance between the positively and negatively charged peptide domains and therefore may inhibit their unwanted interaction. Besides this, the GFLG tetrapeptide used as an enzyme labile spacer may increase the intracellular

release of pentaglutamylated-MTX. The presence of GFLG spacer resulted in very effective penetratin conjugates on MCF-7 cells, but had no effect on the activity of MTX-Glu₅ conjugates on MDA-MB-231 cells. The activity of this conjugate was similar to that of the free MTX. In case of MTX the insertion of the spacer improved the activity of octaarginine conjugate, but the MTX-Glu₅-GFLG-Arg₈ conjugate was fully ineffective. As the cytostatic activity of penetratin (desMet) conjugate was drastically enhanced by the insertion of GFLG spacer on MCF-7 cells, we can suppose that the release of MTX-Glu₅ after cathepsin B cleavage contributed to the higher effect on the activity. The lack of its influence in octaarginine conjugate may mean that the cellular uptake is very inefficient to produce effective MTX-Glu₅ concentration. To support these, the investigation of Cathepsin B enzyme cleavage of conjugates is planned. Based on the results and the inhibitory activity of conjugates and fragments on DHFR enzyme activity the smallest active compounds will be identified. Thus, at the moment it is early to propose even a hypothesis about the mechanism of action and structure-activity relationship. However, we are convinced that the observation presented in this paper is novel and important to communicate.

Our results presented here demonstrate that the delivery of polyglutamylated MTX into tumor cells could result in pronounced improvement on the cytostatic effect on tumor cells *in vitro*. These data also highlight that the choice of cell-penetrating peptide and the design of the conjugate are important, since the selected CPP may have marked influence on the activity. Taking into consideration the possible limitations of CPP in *in vivo* measurements for example degradability, toxicity, further studies are planned to prove the potency of conjugate(s) presented as lead molecules *in vivo*.

5. Conclusions

The resistance of tumor cells against antitumor drugs is a growing problem in the cancer therapy. Therefore new drugs and/or the modification of existing ones circumventing the drug resistance is an intensively researched area. MTX is an effective antitumor compound, but the evolved drug resistance often hampers its use. In many cases the inefficient internalisation or polyglutamation could be responsible for the drug resistance. In this paper we described a new set of conjugates of MTX and its pentaglutamylated derivative (MTX-Glu₅) combined with cell-penetrating peptide – octaarginine and penetratin (desMet). Although both cell-penetrating peptides were successfully used to deliver antitumor drug into cells, as reported in the literature, our results indicate that the penetratin (desMet) conjugates were more effective than the respective octaarginine conjugates under *in vitro* conditions. The delivery of MTX into cells was effective against MTX resistant MDA-MB-231 cells. The penetratin (desMet) conjugates containing MTX-Glu₅ had the same activity on resistant cells than the free MTX on sensitive cells. The MTX-Glu₅-Penetratin (desMet) and MTX-Glu₅-GFLG-Penetratin (desMet) conjugates were very active against resistant tumor cells, and the MTX-Glu₅-GFLG-Penetratin (desMet) had also increased efficacy against sensitive cells. Our results suggest that the delivery of polyglutamylated MTX by cell-penetrating peptide may reopen the way of the effective treatment of tumors. The activity of conjugates also highlights that although the internalisation of cell-penetrating peptide is very similar, the small differences may cause significantly altered activity.

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Appendix A. Supplementary data

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.ejmech.2016.03.034>.

Conflict of interest

All authors declare no conflict of interest.

Author contributions

Designed the experiments: ZB, ISz, GS, FH.

Performed the experiments: ZB, ISz, GS, EO.

Analysed and interpreted the data: ZB, ISz, GS, EO, FH.

Wrote the paper: ZB, ISz, GS, EO, FH.

Revised critically the paper: all authors.

All authors read and approved the final version of the manuscript.

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